

REVIEW WND THEORETICAL STUDIES OF ETHANOL EXTRACT OF *ABRUS* *PRECATORIUS* AND CHARACTERISATION

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Abstract

The objectives of the present study are to analyze qualitative preliminary phytochemical screening and antimicrobial properties of *Abrus precatorius* L. The qualitative preliminary phytochemical performed in aqueous extracts and ethanolic extracts of *Abrus precatorius* L were done and the bioactive compound was identified with TLC plant extract shows three bands for the presence of phenolic compound and R_f values were 0.4, 0.45, and 0.48. The UV-VIS shows the spectra at 278 and 457 nm confirms the organic chromophores and fifty compounds were identified in the ethanolic extracts by GC-MS. The major components present in the *Abrus precatorius* were α -Cyclooctanedione, Furanone, 2,4-Dihydroxy-2,5-dimethyl-3(2H)-furan-3-one, 2-Oxopentanedioic acid, Propenyl Formate, and various other compounds were identified as low level. These phytochemical are responsible for various pharmacological actions. The structure of the identification of ibuprofen sodium using Infra-red spectroscopy were carried various spectral studies. From these studies, i do know about the knowledge of chemistry and spectroscopy. The main objective of this study is to specified and unspecified impurities using high performance liquid chromatography, Assay method of Ibuprofen methods of characterization.

INTRODUCTION

Natural compounds extracted from plants, particularly higher plants, have been suggested as alternative sources for antibiotics. The chemical features of these constituents differ considerably among different species. Because they constitute a potential source of bioactive compounds that have been useful to maintenance of health in humans. Traditionally used medicinal plants produce a variety of compounds of known therapeutic properties. The substances that can either inhibit the growth of pathogens or kill them and have no or least toxicity to host cells are considered for developing new antimicrobial drugs. In recent years, antimicrobial properties of medicinal plants are being increasingly reported from different parts of the world. It is expected that plant extracts showing target sites other than those used by antibiotics will be active against drug-resistant microbial pathogens. Moreover, numerous

plant secondary metabolites such as alkaloids, anthocyanins, flavonoids, quinines, lignins, steroids and terpenoids have found commercial application as drug, dye, flavour, fragrance, insecticide etc., Such fine chemicals are extracted and purified from plant materials by using different solvents. Nowadays most of the secondary metabolite structural diversity is generated by differentially modifying common backbone structures, with the derived compounds having potentially divergent biological activities. Differential modification of common backbone structures can alter the biological activity of a number of plant hormones and secondary metabolites including auxins, glucosinolates, gibberellins and phenylpropanoid derivatives .

In the present investigation *Abrus precatorius* belongs to family fabaceae and used in traditional Ayurvedic medicine, having a important role in the treatment of conjunctivitis in various part of the world. Due to it soothing properties and are expectorant, anti-inflammatory, anti-allergic and anti cancer its leads the present study for phytochemicals analysis in leaves. The obtained phytochemicals were analysed for the TLC,UV-VIS , GC-MS techniques.

MATERIALS AND METHODS:

The ¹H & ¹³C NMR spectra were recorded on a BRUKER 400 MHz NMR spectrometer using DMSO as solvent. the space temperature Fourier transform infrared spectra of 2M5NA were recorded within the range 400-4000 cm⁻¹ at a resolution of ±5 cm⁻¹ employing a BRUKER spectrophotometer equipped with a LiTaO₃ detector, a KBr beam splitter, a He-Ne laser source and a boxcar atomization used for 250 averaged interferograms collected for both the sample and therefore the background. High performance liquid chromatography. an appropriate HPLC instrument equipped with UV detector.

RESULT AND DISCUSSION

Phytochemical Analysis:

Total Impurities : 0.677 % (NMT 1.00%)

Proton NMR spectral analysis

Nuclear magnetic resonance (NMR) spectral analysis is an important analytical technique used to determine the structures of organic compounds. Figure shows that the proton NMR spectra of the molecule NMNPB, exhibits a three singlet peaks were 3.77 & 2.26 ppm (S, 3H, CH₃), 1.32 ppm (S, 3H, CH) due to the presence of methyl group. The secondary amine hydrogen (NH-) signal appeared at 8.91 ppm as a sharp singlet. Two doublet peaks were observed at 7.04 ppm (d, 2H, ArH), 7.76 ppm (d, 2H, ArH), due to the presence of aromatic protons of phenyl ring.. Two triplet peaks were observed at 7.12 (t, 3H, ArH) & 7.53 ppm (t, 3H, ArH) in aromatic ring.

¹³C NMR spectral analysis

Fig.7. shows the ¹³C NMR spectra of Isradipine shows the carbon signals at 167.2 ppm in the downfield respectively due to the highly deshielded carbonyl carbon of the benzamide moiety. The peaks due to aromatic carbons appears at 21.7, 68.4, 120.4, 127.9, 131.1, 112.8, 148.4, 104.1, 150.2, 19.20 ppm in the upfield region is due to methyl carbon.

TLC analysis:

TLC analysis also suggests the presence of different kinds of phytochemicals in leaves extract. Thin layer chromatography was performed on plant extracts using different solvent systems Methanol : Water : Acetone (18:9:1) .

TLC of plant extract in chloroform reports three spots for various phytochemicals. The reported spots are separated with enough space and having various R_f values showing the presence of atleast three phytochemicals in ethanol extracts. In our study, the most suitable TLC system for analysis was shown to be Methanol : Water : Acetone (18:9:1) with the largest discriminating power. Three bands found in this method and its R_f values were 0.4,0.45 and 0.48. This values indicate the presence of phenolic compound.

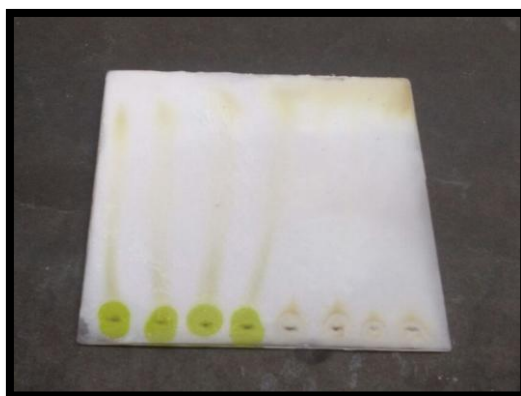


Figure :2 Thin layer cromotography

UV-VIS Analysis

The qualitative UV-VIS profile of ethanolic extract of *Abrusprecatorius* was taken at the wavelength of 300 nm to 800 nm due to the sharpness of the peaks and proper baseline. The profile showed the peaks at 278 and 457 nm with the absorption 4.000, and 1.5088 respectively. Figure 1 shows the absorption spectrum of *Abrusprecatorius* extract and these are almost transparent in the wavelength region of 300-800 nm.

Absorption bands observed pertaining to *Abrusprecatorius* plant extract are displayed in figure 2. In the UV-VIS spectra the appearance of one or more peaks in the region from 200 to 400 nm is a clear indication of the presence of unsaturated groups and heteroatom such as S, N, O. The spectrum for *Abrusprecatorius* extract shows two peaks at positions 278 nm, and 457 nm. This confirms the presence of organic chromophores within the *Abrusprecatorius* extract. Nevertheless, the use of UV-visible spectrophotometry in the analysis of complex media is limited by the inherent difficulties in assigning the absorption peaks to any particular constituents in the system.

Table:3 UV-VIS Analysis of *AbrusprecatoriusL*

S.NO	Wave Length	Absorbance
1	278.00	4.0000
2	457.10	1.5088

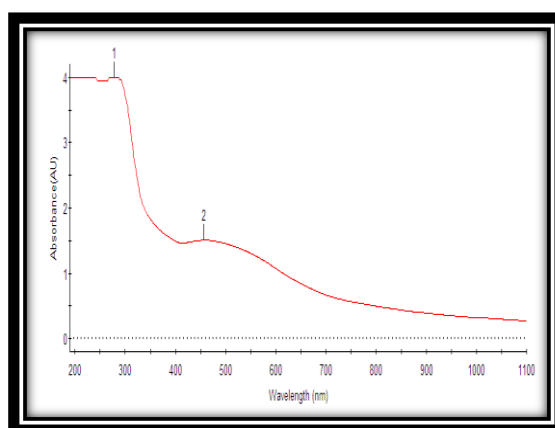


Figure :3 UV-VIS Analysis of *Abrusprecatoriusl*

These absorption bands are characteristic for flavonoids and its derivatives. The flavonoids spectra typically consist of two absorption maxima in the ranges 230-285 nm (band I) and 300-350 nm (band II). The precise position and relative intensities of these maxima give valuable information on the nature of the flavonoids. This is in accordance with the previous literature on *Acorus calamus* (Neha Sahu, Jyoti Saxena 2013)

GC-MS analysis

CONCLUSION:

The Qualitative preliminary Phytochemical performed in aqueous and ethanolic extract of *Abrus precatorius* L were performed. The aqueous extracts showed the presence of coumarin, flavonoids, Tannin, Phenolic compound and quinone and the ethanolic extracts showed coumarin, saponin, terpenoids, flavonoid, tannin, phenolic compound and quinone. The ethanolic extract contains more phytochemical when compared to aqueous extract. In the present study fifty compounds were identified in ethanolic extract by GC-MS. The major components present in the *Abrus precatorius* were α -Cyclooctanedione, Furanone, 2,4-Dihydroxy-2,5-dimethyl-3(2H)-furan-3-one, 2-Oxopentanedioic acid, Propenyl Formate, and various other compounds were identified as low level. These phytochemicals are responsible for various pharmacological actions like antimicrobial and anti-oxidant, anti-inflammation, Anti-cancer, Hepatoprotective, Diuretic, Antiasthma activities.

Reference:

1. Aburjal, Ti Darwish, S. Rmi Alkhalil, A. Mahgzah, A. Al-Abbdi, 2001. Screening of antibiotic resistant inhibitors from local plant materials against two different strains of *Pseudomonas aeruginosa*. J. Ethnopharmacol., 76: 39-44.
2. Adeyeye, A.S., Ogunwale, O.A. and Mofikoya, F.A. (2013) Growth, Dry Matter Accumulation and Shoot Yield of *Celosia argentea* affected by Poultry Manure and Urea Application. International Journal of Agricultural Policy and Research, 1, 210-215.
3. Ajaiyeoba, E.O., P.A. Onocha, O.T. Olarenwaju, 2000. In vitro Anthelmintic properties of *Buchholziacoriacea* and *Gynandropsis gynandra*. J. Pharmaceut. Biol. (in Press)
4. Akinsade, K.A. and D.K. Olukoye, 1995. Vibriocidal activities of some local herbs. J. Diarr. Dis. Res., 13: 127-129.
5. Akinyemi, K.O., A.O. Coker, C. Bayagbon, A.O.B. Oyemfulu, K.A. Ainside and E.O. Omonigbehin, 2000. Antibacterial Screening of Five Nigerian Medicinal Plants Against *S. typhi* and *S. paratyphi*. Journal of the Nigerian infection control association, 3: 1-4.

6. Ashokkumar R, Ramaswamy M. Phytochemical screening by FTIR spectroscopic analysis of leaf extracts of selected Indian medicinal plants. *Int J CurrMicrobiolApplSci* 2014;3(1):395-6.
7. Ayensu, E.S., 2003. *Medicinal Plants of West Africa*. Reference Publication inc. Michigan, U.S.A.
8. Chowdhury, M.N.A., Rahim, M.A., Khalequzzaman, K.M., Humavan, M.R. and Alam, M.M. (2007) Effect of Plant Extracts and Time of Application on Incidence of Anthracnose, Yield and Quality of Mango. *International Journal of Sustainable Crop Production*, 2, 59-68.
9. Dalziel, J.M., 1956. *The useful plants of West Tropical Africa* Crown Agents, London, 612
10. Damodar K. Phytochemical screening, quantitative estimation of total phenolic, flavanoids and antimicrobial evaluation of *Trachyspermum ammi*. *J Atoms and Molecules*, 2011; 1(1): 18
11. Davenport, T.L., Pearce, D.W. and Rood, S.B. (2001) Correlation of Endogenous Gibberellic Acid with Initiation of Mango Shoot Growth. *Journal of Plant Growth and Regulation*, 20, 308-315.
12. Dokoozlian, N.K. (2001) Gibberellic Acid Applied at Bloom Reduces Fruit Set and Improves Size of "Crimson Seed- less" Table Grapes. *HortScience*, 364, 706-709.
13. E. Braunwald, R. D. Bloodwell, L. I. Goldberg, and A. G. Morrow, "Studies on digitals IV observations in man on the effects of digitalis preparations on the contractility of the non-failing heart and on total vascular resistance," *Journal of Clinical Investigation*, vol. 40, no. 1, pp. 52–59, 2010. View at Publisher · View at Google Scholar
14. Gills, L.S., 2011. *Ethno medicinal uses of plants in Nigeria*. Uniben press, University of Benin. Nigeria 9. Hassan, M.M., A.O. Oyewale, J.O. Amupitan, M.S. Abdullahi and E.M. Okonkwo, 2004. Preliminary Photochemical and Antibacterial investigation of crude extracts of the root bark of *Detariummicrocarpum*. *J. Chem. Soc. Nigeria.*, 29(1): 26-29.
15. Ibrahim, D. and H. Osman, 2002. Antimicrobial activity of *Cassia alata* from Malaysia. *J. Ethnopharmacol.*, 45(3): 151-156.
16. Ivanova DG, Singh BR. Nondestructive FTIR monitoring of leaf senescence and elicitor induced changes in plant leaves. *Biopolymers* 2003;72(2):79-85. Available from: <http://www.onlinelibrary.com/doi/10.1002/bip.10297/pdf>.
17. Jayashree VH, Ramesh L. Isolation and identification of a flavone from fruit pulp of *Feronialimonia*. *Int J Curr Pharm Res* 2014;6(4):28-31.

18. K. Khanbabaee and T. van Ree, "Tannins: classification and definition," *Natural Product Reports*, vol. 18, no. 6, pp. 641–649, 2001. View at Publisher · View at Google Scholar · View at Scopus
19. Kumar RS, Moorthy K, Vinodhini R, Punitha T (2013) [Antimicrobial efficacy and phytochemical analysis of Indigoferatritalinn.Afr J Tradit Complement Altern Med 10:518-525.](#)
20. Law-Ogbomo, K.E. and Ekunwe, P.A. (2011) Growth and Herbage Yield of *Celosia argentea* Influenced by Plant Density and NPK Fertilization in Degraded Ultisol. *Tropical and Subtropical Agroecosystems*, 14, 251-260.
21. M. G. L. Hertog, E. J. M. Feskens, P. C. H. Hollman, J. B. Katan, and D. Kromhout, "Dietary antioxidant flavonoids and risk of coronary heart disease: the Zutphen Elderly Study," *The Lancet*, vol. 342, no. 8878, pp. 1007–1011, 1993. View at Publisher · View at Google Scholar
22. M. Z. Barakat, S. K. Shahab, N. Darwin, and E. I. Zahemy, "Determination of ascorbic acid from plants," *Analytical Biochemistry*, vol. 53, pp. 225–245, 1993. View at Google Scholar
23. Marimuthu M, Gurumoorthi P. Phytochemical screening and FTIR studies on wild and common south Indian legumes. *Asian J Pharm Clin Res* 2013;6(2):141-4.
24. Muruganatham S, Anbalagan G, Ramamurthy N. FTIR and SEM-eds comparative analysis of medicinal plants. *Ecliptaalba HASSK and Ecliptaprostrata Linn. Rom J Biophys* 2009;19(4):285-94.
25. N. Kakiuchi, M. Hattori, M. Nishizawa, T. Yamagishi, T. Okuda, and T. Namba, "Studies on dental caries prevention by traditional medicines. VIII. Inhibitory effect of various tannins on glucan synthesis by glucosyltransferase from *Streptococcus mutans*," *Chemical and Pharmaceutical Bulletin*, vol. 34, no. 2, pp. 720–725, 2000. View at Publisher · View at Google Scholar · View at Scopus
26. N. Sheikh, Y. Kumar, A. K. Misra, and L. Pfoze, "Phytochemical screening to validate the ethnobotanical importance of root tubers of *Dioscorea* species of Meghalaya, North East India," *Journal of Medicinal Plants Studies*, vol. 1, no. 6, pp. 62–69, 2013
27. Nascimento, G.G.f., J. Locatelli, Freitas Pc, G.L. Silva, 2000. Antibacterial activity of plant extracts and phytochemicals on antibiotic- resistant bacteria. *Braz. J. microbol.*, 31: 247- 256.

28. Neha Sahu, Jyoti Saxena. Phytochemical Analysis of *Bougainvillea Glabra* Choisy by FTIR and UV -VIS Spectroscopic Analysis, *Int. J. Pharm. Sci. Rev. Res.*, 21(1), Jul – Aug 2013; 96-198
29. Norrie, J. and Keathley, J.P. (2006) Benefits of *Ascophyllum nodosum* Marine plant Extract Applications to Thompson Seedless Grape Production. *ISHS Acta Horticulturae*, 71, 22-28.
30. O. A. Sodipo, J. A. Akiniyi, and U. S. Ogunbano, “Studies on certain characteristics of extracts of bark of *Pausinystalia johimbe* and *Pausinystalia macroceras* (K.Schum.) Pierre ex Beille,” *Global Journal of Pure and Applied Sciences*, vol. 6, no. 1, pp. 83–87, 2000
31. Olowosuluand, A.K. And Ibrahim, Y.K.E. (2006) Studies on the Antimicrobial Screening of Aqueous Extracts of Five Plants Used in Folk Medicine in Nigeria. *West African Journal of Biological Science*, 3, 21-26.
32. Omode AA, Fatoki OS and Olaogun KA. Phytochemical properties of some under exploited and non - conventional oil seeds. *J. Agric. Food Chem.* 1995; 43(11): 2850 - 2853
33. Osuagwu, G.G.E. and Ibeabuchi, I.C. (2010) The Influence of Aqueous Leaf and Stem Extracts of *Adenialobata* (Jacq) on the Flowering and Fruiting of Okra (*Abelmoschus esculenta*) and Groundnut (*Arachis hypogea*). *African Journal of Biotechnology*, 9, 3260-3263.
34. S. Y. Kim, J. H. Kim, S. K. Kim, M. J. Oh, and M. Y. Jung, “Antioxidant activities of selected oriental herb extracts,” *Journal of the American Oil Chemists' Society*, vol. 71, no. 6, pp. 633–640, 2000. View at Publisher · View at Google Scholar · View at Scopus
35. Stehfest K, Toepel J, Wilhelm C. The application of micro-FTIR spectroscopy to analyze nutrient stress-related changes in biomass composition of phytoplankton algae. *Plant Physiol Biochem* 2005;43(7):717-26.
36. Sundaram S, Palanisamy CP, Velliyur KG. Chromatographic and spectrophotometric analysis of bioactive compounds from *Cayratia trifolia* (L) stem. *Int J Pharm PharmSci* 2016;8(6):56-64.
37. Tiwari, P., Kumar, B. (2011) Phytochemical screening and Extraction: A Review, *Journal of Internationale Pharmaceutica Scientia*, 1:98-106.
38. Y. Kashiwada, L. Huang, R. E. Kilkuskie, A. J. Bodner, and K.-H. Lee, “New hexahydroxydiphenyl derivatives as potent inhibitors of HIV replication in H9 lymphocytes,” *Bioorganic and Medicinal Chemistry Letters*, vol. 2, no. 3, pp. 235–238, 1992. View at Publisher · View at Google Scholar · View at Scopus

39. Yang J, Yen HE. Early salt stress effects on the changes in chemical composition in leaves of ice plant and Arabidopsis. A Fourier transform infrared spectroscopy study. *Plant Physiol* 2002;130(2):1032-42.