

NANOTECHNOLOGY METHODS FOR DNA ISOLATION

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Abstract

Deoxyribonucleic acid (DNA) is unique to each individual and will remain unchanged for a lifetime as it follows the rules of the Mendelian lineage. However DNA analysis is a widely accepted technique for individual identification. Extracting and isolating DNA from ancient bone samples and mutilated body parts is an even more difficult and challenging task for the forensic scientist. But newer technology, such as nanotechnology, will help to analyze such models for DNA. Therefore, in this research paper we have shortlisted some research papers that used different nanoparticles to extract DNA. The different nanoparticles used for DNA extraction were cited and briefly explained.

Keywords: Nanotechnology; Personal Identification; DNA; Forensic; Nanoparticles; Copper nanoparticles; Gold nanoparticles; Magnetic Nanoparticles

Introduction

Personal Identification Using DNA

Traditionally, human remains have been identified individually or forensically based on fingerprint, otontology or bone evidence, facial features, scars, marks or other special features. In many cases, available methods have not proven effective because the amount of destruction or destruction of debris or debris is very high [1].

Disasters terrorist attacks, traffic accidents, wars, fires, explosions, plane crashes highly, decomposed body or severely burnt and other harrowing events individual identification or forensic identification become very tough for the investigation man from skeletal remains [2-4].

During decomposition human body undergoes a series of multiple changes and the rate of decomposition may vary due to factors like monsoon, microorganisms growth, and insect and wild scavengers [1]. During this time the soft tissues are often destroyed and may have no dental history [2]. Some of conditions Deoxyribo Nucleic Acid (DNA) can prove to be the most very very essential and useful source of identification or a good another source for identification purposes [2]. Deoxyribonucleic acid (DNA) is responsible for genetic inheritance, So the discovery of the dual helix structure of DNA in 1953 had a major impact And in all fields of science, including forensic investigations Led to significant changes.

This understanding is the foundation for the development of new techniques and methods

It is based on the individual DNA sequence.

DNA is made up of phosphate spines, deoxyribose sugars and nitrogen bases Where phosphate backbone and deoxyribose sugar remain unchanged in all individuals, Only nitrogen bases are different for each person. Thus, human DNA makes up 99.9 percent of the scenery per person , Only 0.1 percent are unique to each person, though the possibility of having same genetic

material is about approximately 10-15 between two mismatched individual [5,6]. These minute differences can be detected in human DNA sequences and based on these differences between the 1984 DNA sequences The first forensic DNA tests were performed [7]. In recent years, it has become widespread that DNA can lead to the personal identification of human remains, I.e. bone residues in most cases. DNA sample identification and specification play a very important role in mass disaster And also helps identify suspects in civil and criminal cases. But at the same time, many organisms exposed to microorganisms, moisture and bone residues Or it should be noted that the decomposition of genetic material by mineral compounds reduces DNA levels and the reliability of extracting good quality DNA. The use of small-scale loci multiplication by polymerase chain reaction (PCR) to overcome such a problem yields excellent results for further analysis of genetic material. However, in forensic examination of skeletons, in the context of individual identification, PCR-ready genetic material extraction has become an important and challenging task. For forensic uses, DNA is usually separated from many biological evidence, such as blood, saliva, semen, hair, urine, skin, tissues, teeth, and bones [8-20].

From all of these bone and dental specimens, especially from the ancient / buried / autopsy bone remains specimens, the examination is a chalky endeavor as the skeletal remains bring problems to the DNA extraction process. Effective disruption of nucleoprotein / protein-DNA complexes in cells and dinosaur embedded cells. In addition, even pulse-sized bone dust does not have uniform consistency, usually due to low levels of cellular material and age-related degradation of DNA in the bone and polymerase chain reaction (PCR) inhibitors Integrating. In addition to DNA hydrolysis, DNA molecules undergo biochemical degradation such as hydrolytic and oxido-dive damage [21,22] by cellular nuclei and microbial dextro-dione.

There are several commercial products for DNA isolation, including column chromatography or extraction following agarose gel electrophoresis, But these techniques fail to avoid contamination or complicated procedures. It is necessary to get rid of any contaminants such as DNA protein and RNA isolated from biological samples. The process of genetic DNA isolation and purification has progressed significantly over the past decade,

But improvements are needed in the mechanisms for DNA isolation. There is a need to replace old labor and time consuming traditional techniques,

It generally relies on the use of the phenol-chloroform method. All of these DNA extraction techniques are time consuming, multi-step and use for organic solvent extraction, alcohol precipitation and centrifugation [23]. However, nanotechnology has recently entered into DNA isolation to answer and deal with such issues.

Forensic Nanotechnology

The advent of nanotechnology controls atoms and molecules individually, modifying and making them usable with extraordinary levels of precision. The word 'nano' is derived from the Greek word for dwarf, which refers to one billion (10^{-9}) nanometers (nm) [24]. It is associated with growing material or devices with a size of 100 nm or less. Nanoparticles are applied in a variety of fields, including electronics, engineering, physics, and semantics, and have found application in the field of medical science [25]. Advances in nanotechnology have led to the development of new methods that are innovative and have advanced properties that demonstrate their validity in all fields of science and technology. Nanotechnology currently plays an important role in the fields of biology and biology [26,27].

Currently, nanoparticles have attracted great attention for their applications in biochemistry, medicine and biotechnology due to their unique size and physical properties. In the case of biodegradation and refining, the viable solution is to replace the nanoparticles with nanoparticles because they are more proportional to the surface. Integrated systems have been developed to isolate DNA, which use solid-phase support Integrated Systems Have Been Developed To Isolate Dana, With Us Us Solita-Pache Support Magnetic particles have been used successfully to extract genetic DNA from body fluids such as blood, saliva and semen [30]. Magnetic particles coated with different polymers such as agros, carboxyl and silica have been studied for DNA isolation [31].

Solid phase systems are used to purify DNA-silica-based particles, glass fibers, anion-transfer carriers, and modified magnetic beads because they have the advantage of minimal DNA decay during refining [32-41]. Extraction of DNA from blood was done by ZM Saeed et al using magnetic nanoparticles. [40], Rui Kong et al. Use magnetic microspheres. [41] and ZM Syed et al. [42], from cell culture using magnetic nanoparticles by ZM Saeed et al. [42] and Zhang Zhao et al. Using magnetic nanoparticles coated with salicylic acid. [43], A. Sebastianelli et al. From soil using silica-magnetic nanoparticles. [44], Ji Shan and others using magnetic nanoparticles from urine. [45] and Li Yi et al. From saliva using magnetic nanoparticles. [46]. The phenol / chloroform method uses toxicity, timing and multi-step and organic solvent extraction, alcohol precipitation and centrifugation [47]. The method has a limit when dealing with small amounts of DNA. The use of magnetic carrier technology to separate living cells, such as DNA, RNA, and proteins [48,49], has become a widespread technique.

Lotha et al. [50] The unique property of copper nanoparticles was first used to isolate DNA from bone residues. Copper nanoparticles are synthesized in the microwave using diethylene glycol (DEG), ascorbic acid and polyvinyl pyrrolidone (PVP), which bind highly to DNA. The novelty of this method is that copper nanoparticles have not yet been used for DNA isolation.

Furthermore, the author said that this developed method could be easily used to isolate DNA from bone remains.

Mundaha et al. [51] DNA quality and quantity improved when DNA was extracted from gram-negative bacteria by extraction methods (boiling, alkaline lysis, and salt extraction) in the presence of ZnO nanoparticles. The resulting heat effectively caused the pathogen to decompose. The resulting DNA is extracted from the cell body and transferred to the PCR system. This led to a step-by-step demonstration of the success of the real-time PCR system.

Conclusion

The use of nanotechnology to identify unknown human remains using forensic genetics / DNA has the potential to provide accurate personal identification, even from small, dangerous, trace samples. Forensic analysis needs to increase DNA extraction capabilities because sample sizes are often limited and may introduce incomplete DNA extraction dependencies, thus altering the interpretation of the results. Reproduction and nanomaterials used in the extraction steps require research. None of the standardized extraction methods currently used in forensic laboratories are used because of the DNA extraction protocols based on different types of nanoparticles. It may limit the ability to compare forensic models in different forensic research and laboratory groups, countries, or case studies. Finally, laboratory standardization based on nanoparticles in chip DNA forensic methods should be explored. When these problems are solved, forensic scientists

will be able to combine nanomaterials and DNA analysis into a commonly used casework, using nanotechnology to provide robust, reliable sources for personal identities.

References

1. LC Richard (2009) Thesis University of Indiana University Purdue.
2. J Ye, Ji EJ, Parrg X, Zheng C, Jiang, et al. (2004) *Tu J Forensic Sci* 49 : 754759.
3. MM Hasan, T Hossain, AK Majumder, P Momtaz, T Sharmin, et al. (2014) *Dhaka Univ J Biol Sci* (23): 101-107.
4. J Jakubowska, A Maciejewska, R Pawiowski (2012) Comparison of three methods of DNA extraction from human bones with different degrees of degradation. *Int J Legal Med* 126(1): 173-178.
5. DA Schultz (2009) *The Encyclopedia of American Law*. Infobase Publishing.
6. RE Gaensslen, HA Harris, HC Lee (2007) *Introduction to Forensics & Criminalistics*. Mc Graw Hill Companies Inc, USA.
7. MA Luftig, S Richey (2001) *New England law review* 35: 609-613.
8. S Ghatak, RB Muthukumar, SK Nachimuthu (2013) A Simple Method of Genomic DNA Extraction from Human Samples for PCR-RFLP Analysis. *Journal of Biomolecular Techniques* 24(4): 224-231.
9. DK Lahiri, B Schnabel (1993) DNA isolation by a rapid method from human blood samples: Effects of MgCl₂, EDTA, storage time, and temperature on DNA yield and quality. *Biochemical Genetics* 31: 321328.
10. RW Cone, ML Huang, R Ashley, L Corey, (1993) Human herpesvirus 6 DNA in peripheral blood cells and saliva from immunocompetent individuals. *J Clin Microbiol* 31(5): 1262-1267.
11. DJ Walsh, AC Corey, RW Cotton, L Forman, G L Herrin, et al. (1992) Isolation of deoxyribonucleic acid (DNA) from saliva and forensic science samples containing saliva. *J Forensic Sci* 37: 387-395.
12. NW Schwark, V Malyusz, H Fremdt, C Koch, T Schwark (2006) *International Congress Series* 1288: 565-567.
13. J Griffin (2013) Methods of sperm DNA extraction for genetic and epigenetic studies. *Methods Mol Biol* 927: 379-384.
14. Z Guan, Y Zhou, J Liu, X Jiang, S Li (2013) A Simple Method to Extract DNA from Hair Shafts Using Enzymatic Laundry Powder *PLoS ONE* 8: e69588.
15. LE Bali, A Diman, A Bernard, NHC Roosens, SCJD Keersmaecker (2014) Comparative study of seven commercial kits for human DNA extraction from urine samples suitable for DNA biomarker-based public health studies. *Biomol Tech* 25(4): 96-110.
16. JV Sidorova, BV Biderman, EE Nikulina, AB Sudarikov (2012) A simple and efficient method for DNA extraction from skin and paraffin- embedded tissues applicable to T-cell clonality assays. *Exp Dermatol* 21(1): 57-60.
17. JMS Bartlett, D Stirling(2003) A short history of the polymerase chain reaction. *PCR Protocols Methods in Molecular Biology*, Human Press 226: 33-34.
18. R Gaytmenn, D Sweet (2003) *J Forensic Sci* 48: 622-625.

19. Z Presecki, H Brkic, D Primorac, I Drmic (2000) Perception of Forensic Odontology and its Practice among the Local Dentists of an Institution . *ActaStomatol Croat* 34: 21-24.
20. PV Mandrekar, L Flanagan, A Tereba (2002) Improved performance for forensic casework: Extraction and isolation updates for the Maxwell 1 16 instrument Profiles in DNA. 5: 11-13.
21. T Lindahl(1993) Instability and decay of the primary structure of DNA. *Nature* 362: 709-715.
22. JG Shewale, RH Liu (2013) *Forensic DNA Analysis: Current Practices and Emerging Technologies*, CRC Press, Taylor and Francis, USA.
23. A Bandyopadhyay, S Chatterjee, K Sarkar (2011) *Current science* 101: 210-214.
24. V Labhasetwar, DL Pelecky (2007) *Biomedical Applications of Nanotechnology*. Wiley, New York, USA.
25. OV Salata (2004) *Journal of Nanobiotechnology* 2: 1-6.
26. JH Min, MK Woo, HY Yoon, J WJang, JH Wu (2014) Isolation of DNA using magnetic nanoparticles coated with dimercaptosuccinic acid. *Analytical Biochemistry* 447: 114-118.
27. GK Kouassi, J Irudayaraj (2006) Magnetic and Gold-Coated Magnetic Nanoparticles as a DNA Sensor. *Anal Chem* 78: 3234-3241.
28. Z Shan, Z Zhou, H Chen, Z Zhang, Y Zhou, et al. (2012) *Journal of Chromatography B* (881-882): 63- 68.
29. R Boom, CJA Sol, MMM Salimans, CL Jansen, PME Wertheim van Dillen, et al. (1990) Rapid and simple method for purification of nucleic acids. *J Clin Microb* 28: 495-503.?
30. KA Melzak, CS Sherwood, RFB Turner, Ch A Haynes (1996) Adsorption and elution characteristics of nucleic acids on silica surfaces and their use in designing a miniaturized purification unit. *J Colloid Interface Sci* 181: 635-644.
31. H Tian, AFR Huhmer, JP Landers (2000) *Anal Biochem* 283: 175-191.
32. MC Breadmore, KA Wolfe, IG Arcibal, WK Leung, D Dickson, et al. (2003) *Anal Bioche* 75: 1880-1886.
33. GNM Ferreira, JMS Cabral, DMF Prazeres (2000) *Biotechnol Prog* 16: 416-424.
34. E Thwaites, SC Burton, A Lyddiatt, J Chromatogr (2002) 943: 77-90.
35. HN Endres, JAC Johnson, CA Ross, JK Welp (2003) *Biotechnol Appl Biochem* 37: 259-266.
36. MA Teeters, SE Conrardy, BL Thomas, TW Root, EN Lightfoot (2003) Adsorptive membrane chromatography for purification of plasmid DNA. 989: 165-173.
37. PR Levison, SE Badger, J Dennis, P Hathi, MJ Davies (1998) 816: 107-111.
38. PR Levison, SE Badger, P Hathi, MJ Davies, IJ Bruce (1998) New approaches to the isolation of DNA by ion-exchange chromatography. 827(2): 337-344.
39. J Prodelalova, B Rittich, A Spanova, K Petrova, MJ Benes (2004) 1056: 43-48.
40. ZM Saiyed, CN Ramchand (2007) Extraction of Genomic DNA Using Magnetic Nanoparticles (Fe₃O₄) as a Solid-Phase Support. *Am J Infect Dis* 3(4): 225-229.
41. R Gong, L Shengying (2014) *Int J Nanomedicine* Extraction of human genomic DNA from whole blood using a magnetic microsphere method 9: 3781-3789.
42. ZM Saiyed, CN Ramchandand, SD Telang (2008) *J Phys Condens Matter* 20: 1-5.
43. Z Zhou, US Kadam, J Irudayaraj (2013) One-stop genomic DNA extraction by salicylic acid-coated magnetic nanoparticles. *Anal Biochem* 442(2): 249-252.
44. A Sebastianelli, T Sen, IJ Bruce (2008) Extraction of DNA from soil using nanoparticles by magnetic bioseparation. *Lett ApplMicrobiol* 46(4): 488-491.
45. Z Shana, Z Zhou, H Chen, Z Zhang, Y Zhou, (2012) *J Chromatogr B* (881882): 63-68.
46. L Yi, Y Huang, T Wu, J Wu (2013) A magnetic nanoparticles-based method for DNA extraction from the saliva of stroke patients. *Neural Regen Res* 8(32): 3036-3046.

47. J Sambrook, D Russell (2001) *Molecular Cloning: a Laboratory Manual* (3rd edn); Cold Spring Harbor NY Cold Spring Harbor Laboratory.
48. PJ Robinson, P Dunnill, MD Lilly (1973) The properties of magnetic supports in relation to immobilized enzyme reactors. *Biotechnol Bioeng* 15(3): 603-606.
49. M Magnani, L Galluzzi IJ Bruce (2006) The use of magnetic nanoparticles in the development of new molecular detection systems. *J Nanosci Nanotechnol* 6: 2302-2311.
50. A Lodha, A Pandya, RK Shukla (2016) *Nanotechnology: An Applied and Robust Approach for Forensic Investigation*. *Forensic Res CriminolInt J* 2(1): 00044.
51. RA Muntaha, Majid HA, Ayad MA, Shaymaa RA (2015) *Intl Conf on J Medical Genetics, Cellular & Molecular Biology, Pharmaceutical & Food Sciences Istanbul*.
52. KH Cheong, DK Yi, JG Lee, JM Park, MJ Kim (2008) Gold nanoparticles for one step DNA extraction and real-time PCR of pathogens in a single chamber. *Lab Chip* 8(5): 810-813.

